Serum Iron Markers Are Inadequate for Guiding Iron Repletion in Chronic Kidney Disease

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Summary
Background and objectives Iron (Fe) overload may complicate parenteral Fe therapy used to enhance the efficacy of erythropoietic-stimulating agents in the treatment of anemia of chronic kidney disease. However, serum Fe markers are influenced by inflammation or malignancy and may not accurately reflect the amount of body Fe.

Design, setting, participants, & measurements We studied the relationship between parenteral Fe therapy, conventional serum Fe markers, and liver iron concentration (LIC) measured using magnetic resonance R2 relaxometry (FerriScan) in 25 Fe-deficient predialysis chronic kidney disease patients before and 2 and 12 weeks after single high-dose intravenous Fe and in 15 chronic hemodialysis patients with elevated serum ferritin (>500 μg/L).

Results In predialysis patients, there was strong dose dependency between the administered Fe dose and changes in LIC at weeks 2 and 12; however, no dose dependency between Fe dose and changes in ferritin or transferrin saturation (TSAT) were observed. In hemodialysis patients, LIC correlated with the cumulative Fe dose and duration of dialysis but not with current ferritin or TSAT. The cumulative Fe dose remained a significant independent predictor of LIC in a multiple regression model. Two dialysis patients who received >6 g parenteral Fe had substantially elevated LIC >130 μmol/g, which is associated with hemochromatosis.

Conclusions In Fe-deficient predialysis patients, intravenous Fe therapy is associated with increases in LIC unrelated to changes in conventional Fe markers. In hemodialysis patients, TSAT and ferritin are poor indicators of body Fe load, and some patients have LICs similar to those found in hemochromatosis.

Introduction Chronic kidney disease (CKD) is commonly accompanied by the development of anemia that is characterized by poor intestinal iron (Fe) absorption, low ferritin levels, and requirement for parenteral Fe supplementation (1–3). There is good evidence that intravenous Fe therapy should be administered as a standard therapy for Fe deficiency (ferritin <100 μg/L, transferrin saturation [TSAT] <20%) in conjunction with or before therapy with erythropoietic-stimulating agents in anemia of CKD and to maintain adequate Fe stores in dialysis patients (4,5). Hemoglobin and ferritin concentrations have been shown to increase significantly in CKD patients after intravenous Fe compared with oral Fe therapy (6). For ease and convenience, intravenous Fe 500 to 1500 mg can be administered over a 4-hour period (7). Dialysis patients regularly require 50 to 200 mg monthly of intravenous Fe to maintain their Fe stores (8,9). Although it is generally assumed that restoration of hemoglobin toward the target range is a good outcome of this therapy, it is well known that Fe overload and Fe toxicity may be adverse consequences of this therapy. Both the American and Australian guidelines (4,5) recommend caution with the routine administration of intravenous Fe if the serum ferritin is >500 μg/L. However, the upper limit of a safe serum ferritin level remains unresolved (10). Serum ferritin levels between 300 and 1200 μg/L are associated with the lowest mortality risk after adjusting for malnutrition and inflammation (11), and only serum ferritin levels >2000 μg/L have been associated with hemochromatosis in dialysis patients (3). Unfortunately, serum Fe markers do not accurately reflect the amount of Fe in the body, because factors such as infections, inflammatory diseases, or malignancy can alter ferritin levels in the body. Thus, some authors have suggested that no specific level of serum ferritin should be stated as an upper limit for Fe treatment (10).

Recently, a noninvasive magnetic resonance imaging (MRI)-based R2 relaxometry method (FerriScan) was developed that accurately measures liver Fe concentration (LIC) in a range of diseases (12). LICs >130 μmol/g dry weight are associated with increased risks of liver injury (13), whereas cardiac toxicity occurs when LIC exceeds 270 μmol/g (14). Liver biopsy
is rarely performed in subjects with CKD, and retrospective quantitative phlebotomy as a direct measure of total body Fe burden is not possible because of the anemic status of the patients and their consequent limited tolerance of phlebotomy. Given the uncertainties surrounding the accuracy of serum Fe biochemistry as a direct measure of Fe status, R2 relaxometry presents a readily available and validated noninvasive method of measuring LICs in vivo to assess the relationship between conventional biochemical markers of Fe stores, intravenous Fe therapy, and liver Fe load in CKD patients. The aims of this study were to (1) prospectively evaluate the effects of a single high-dose Fe infusion on LIC in Fe-deficient CKD stage 3 to 5 patients who had previously not received parenteral Fe and (2) characterize the extent of hepatic Fe overload in hemodialysis patients receiving regular intravenous Fe replacement.

Materials and Methods

Prospective Study in Iron-Deficient CKD Subjects

The primary objective of this substudy was to prospectively assess LIC before and after a single high-dose intravenous Fe infusion and to compare LIC with conventional markers of Fe stores. This cohort comprised patients with stage 3 to 5 CKD (estimated GFR, 10–60 ml/min per 1.73 m2), who were scheduled to receive 10 to 20 mg/kg of intravenous Fe-poly maltose (Ferrosig, Sigma Pharmaceuticals, Croydon, Australia) if they had (1) anemia, using World Health Organization definitions of hemoglobin <120 g/L for women and <130 g/L for men (15), (2) Fe deficiency (TSAT < 20% and/or serum ferritin < 100 µg/L), (3) not previously received parenteral Fe, and (4) no contraindications to MRI. The first MRI assessment of LIC was scheduled on the day of intravenous Fe therapy just before infusion; postinfusion scans and biochemical studies were obtained at weeks 2 and 12. The primary endpoint was calculated as the change in mean LIC from baseline to follow-up. A sample size of 20 was expected to provide 80% power to detect a change in mean LIC over time of 7.7 µmol/g dry weight, equating to an effect size of 66%, using a paired t test with a two-tailed α level of 0.05 (16). This was based on an SD of LIC of 11.7 µmol/g reported in hemodialysis patients (17).

Cross-Sectional Study in Hemodialysis Subjects

The primary objective of this substudy was to assess whether LIC in hemodialysis patients with serum ferritin levels above the upper recommended guidelines was best predicted by ferritin, TSAT, or cumulative Fe dose. This cohort comprised chronic hemodialysis patients treated between September 2009 and February 2010. The inclusion criteria were (1) maintenance hemodialysis of ≥12 months, (2) native arteriovenous fistula, (3) parenteral Fe therapy (Ferrosig, usually 50 to 200 mg monthly) over the past 12 months, (4) serum ferritin >500 µg/L on two consecutive occasions 3 months apart, (5) full history of total parenteral Fe treatment since start of dialysis, (6) alcohol consumption of less than two standard drinks per day, and (7) exclusion of significant liver disease. Serologic testing was used to exclude chronic viral hepatitis B or C infection. Serum Fe markers and LIC-R2 relaxometry were measured at least 2 weeks after the last intravenous Fe dose. These studies were approved by the Hospital Ethics Committee, and all subjects consented to participation in the study.

MRI

MRI was conducted on a 1.5-T whole body imaging unit (Siemens MAGNETOM Vision Plus) using a recently described method (12). This method measures the mean liver proton transverse relaxation rate (R2), which has a high sensitivity and specificity for prediction of LIC (12). Images were acquired in partial Fourier mode with a multislice single-spin-echo pulse sequence, with the pulse repetition time = 2500 ms, spin echo times = 6, 9, 12, 15, and 18 ms, and slice thickness = 5 mm. All spin-echo sequences were performed using fixed gain control. Subjects were scanned together with a bag of normal saline, which acted as a long T2 standard to correct for any drift between different image acquisitions. Subjects were positioned to locate the liver central to the chest coil. Eleven slices were collected, with the gap between slices being 5 mm. R2 values were calculated throughout the largest cross-section of the liver by curve fitting the equation for the bi-exponential decay in transverse magnetization after an single-spin-echo pulse sequence to the voxel intensity data as a function of echo time with radiofrequency field intensity-weighted spin density projection (18). Mean LIC was calculated from mean R2 values as previously reported (12). The accepted normal range of LIC in healthy subjects is <30 µmol/g (Figure 1) (19). Serum Fe and ferritin concentrations and TSAT were measured or derived using standard methods.

Statistical Analyses

Statistical analysis was performed with Systat software package version 10 (SPSS, Chicago, IL). Results are presented as mean ± SD for continuous variables and number and percentage for categorical variables. Differences between subjects were tested by nonparametric Mann-Whitney U test statistic for continuous data and Fisher exact test for categorical data. ANOVA was used to compare within-group changes over time in the first substudy. Non-CKD patients with secondary Fe overload and LIC >60 µmol/g are considered eligible for chelation therapy, whereas those with LIC >130 µmol/g are at increased risk of liver injury (13). Thus, these values were used as threshold levels for determination of the number of dialysis patients above and below these limits. All P values quoted are two tailed.

Results

Prospective Study in Iron-Deficient CKD Subjects

Twenty-five CKD patients (stage 3, n = 6; stage 4, n = 14; stage 5, n = 5) qualified for intravenous Fe
infusion according to the hospital protocol and com-
completed the three follow-up visits. Baseline char-
acteristics of these patients are summarized in Table 1. The
dose of intravenous Fe ranged from 1000 to 1500 mg,
and the average Fe dose per body weight was 15 ± 3
g/kg. At week 2, the hemoglobin averaged 113 ± 12
g/L ($P$ = not significant versus baseline), TSAT aver-
age 31 ± 12% ($P < 0.00001$ versus baseline), and

Figure 1. Liver R2 images and distributions for four subjects with different degrees of iron (Fe) overload and pathologic conditions: (A) healthy control, (B) hereditary hemochromatosis, (C) ESKD patient with >6 g cumulative Fe dose, and (D) Fe deficient CKD patient before and 2 and 12 weeks after 1 g intravenous Fe (top to bottom). Note that the liver R2 images are superimposed on standard spin-echo images for registration purposes. Note that to enable visualization of the heterogeneity of R2 within each liver, the color scale within each liver is adjusted for each image such that zero corresponds to voxel R2 of zero, whereas the maximum of the color scale is scaled to the maximum R2 value within the liver.

### Table 1. Baseline characteristics of the iron-deficient CKD cohort and the hyperferritinemic hemodialysis cohort

<table>
<thead>
<tr>
<th></th>
<th>CKD</th>
<th>HD</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>25</td>
<td>15</td>
</tr>
<tr>
<td>Age (years)</td>
<td>65 ± 15</td>
<td>61 ± 12</td>
</tr>
<tr>
<td>Gender (male/female)</td>
<td>17/8</td>
<td>10/5</td>
</tr>
<tr>
<td>Serum creatinine (mg/L)</td>
<td>296 ± 130</td>
<td>—</td>
</tr>
<tr>
<td>Hemoglobin (g/L)</td>
<td>107 ± 8</td>
<td>116 ± 9</td>
</tr>
<tr>
<td>Transferrin saturation (%)</td>
<td>15 ± 6</td>
<td>31 ± 10</td>
</tr>
<tr>
<td>Ferritin (mg/L)</td>
<td>67 ± 56</td>
<td>782 ± 170</td>
</tr>
<tr>
<td>CRP (mg/L)</td>
<td>1.2 ± 2.6</td>
<td>4.9 ± 4.1</td>
</tr>
<tr>
<td>Transfusions (N)</td>
<td>0</td>
<td>3.8 ± 2.7</td>
</tr>
<tr>
<td>Time on dialysis (days)</td>
<td>—</td>
<td>899 ± 353</td>
</tr>
<tr>
<td>Cumulative Fe dose (mg)</td>
<td>—</td>
<td>6560 ± 3098</td>
</tr>
<tr>
<td>Mean monthly Fe dose (mg)</td>
<td>—</td>
<td>217 ± 87</td>
</tr>
<tr>
<td>Weekly erythropoietin dose (U/wk)</td>
<td>—</td>
<td>7870 ± 4360</td>
</tr>
<tr>
<td>Liver Fe concentration (μmol/g)</td>
<td>20.6 ± 7.9</td>
<td>81.2 ± 58.3</td>
</tr>
</tbody>
</table>
ferritin averaged $563 \pm 282 \, \mu g/L$ ($P < 0.0001$ versus baseline), and at week 12, the mean hemoglobin was $120 \pm 13 \, g/L$ ($P < 0.00001$ versus baseline), TSAT was $25 \pm 9\%$ ($P < 0.0001$ versus baseline), and ferritin was $299 \pm 221 \, \mu g/L$ ($P < 0.001$ versus baseline; Figure 2). LIC increased significantly to $46.1 \pm 15.6 \, \mu mol/g$ at week 2 and $33.7 \pm 11.3 \, \mu mol/g$ at week 12 (Figure 2). A comparison of LIC at week 2 in subjects receiving Fe amounts below or above the median dose showed that LIC remained $\leq 30 \, \mu mol/g$ in 25% of subjects given $<14.5 \, mg/kg$ Fe, whereas all subjects receiving $\geq 14.5 \, mg/kg$ of Fe had LIC $>30 \, \mu mol/g$. The mean increase in LIC from baseline was $25.4 \pm 15.6 \, \mu mol/g$ (152 ± 126%) at week 2 and $13.0 \pm 11.2 \, \mu mol/g$ (80 ± 89%) at week 12. The change in TSAT from baseline tended to show a dose dependency at week 2 but not at week 12, whereas there was no dose dependency for changes in serum ferritin either at week 2 or 12. The increase in LIC showed a clear dependence on the administered Fe dose at both weeks 2 and 12 (Figure 3).

Cross-Sectional Study in Hemodialysis Subjects

Twenty patients were screened to participate in the study, but five failed to undergo $R_2$ relaxometry (claustrophobia, $n = 1$; pacemaker, $n = 1$; canceled two appointments, $n = 2$; malignancy, $n = 1$). Patients were dialyzed three times weekly for $272 \pm 24$ minutes on a Fresenius 4008 machine using FX80 high-flux dialyzers at an average blood flow of $329 \pm 25 \, ml/min$. Baseline characteristics of these patients are summarized in Table 1. LIC correlated with the cumulative Fe dose ($R^2 = 0.44$, $P < 0.01$) and the duration of dialysis ($R^2 = 0.39$, $P < 0.05$) but not with current ferritin or TSAT (Figure 4). The cumulative Fe dose remained a significant independent predictor of LIC ($R^2 = 0.69$, $P < 0.05$) in a multiple regression model that included C-reactive protein (CRP), ferritin, TSAT, current and cumulative Fe dose, and duration of dialysis. Nine patients (60%) had LIC $>60 \, \mu mol/g$. All seven subjects with $\geq 6000 \, mg$ cumulative Fe dose had LIC $>60 \, \mu mol/g$ compared with only two of eight subjects with cumulative Fe dose $<6000 \, mg$. Two of seven subjects with $\geq 6000 \, mg$ cumulative Fe dose had LIC $>130 \, \mu mol/g$ (Figure 1).
and all but one subject had serum ferritin significant, although all subjects had TSAT and serum ferritin compared with baseline were still intravenous Fe (23). At 12 weeks, changes in TSAT may take up to 14 days to reach steady state after reflect Fe overload, because changes in Fe indicators in TSAT. However, these changes are not expected to persist Fe overload, because changes in Fe indicators may take up to 14 days to reach steady state after intravenous Fe (23). At 12 weeks, changes in TSAT and serum ferritin compared with baseline were still significant, although all subjects had TSAT <50%, and all but one subject had serum ferritin <500 µg/L as recommended by guidelines (4,5). On the other hand, 56% of the predialysis CKD subjects had LIC in excess of the upper limit of normal (<30 µmol/g) 12 weeks after high-dose parenteral Fe, although none had levels >60 µmol/g. In our cohort, only changes in LIC, but not serum ferritin or TSAT, showed a dose dependency to the administered Fe dose. These findings suggest that, in Fe-deficient CKD patients, a single, high-dose Fe infusion only leads to a transient liver Fe loading that is dependent on the amount of Fe administered. Although increases in LIC in predialysis CKD patients after Fe infusion are of smaller magnitude and transient, and thus seem to be safe, repeated infusions over intervals <12 weeks could result in significant Fe load.

In the second substudy, we deliberately selected CKD patients on dialysis with serum ferritin levels above what is considered to be the safety threshold (4,5). Thus, one would expect the majority of these patients to show significantly increased LIC and serum ferritin to predict LIC. Our findings suggest that this may occur only in hemodialysis patients with persistently increased serum ferritin levels who have received considerable amounts of parenteral Fe. Interestingly, we found no correlation between serum Fe markers and LIC, but a strong correlation between LIC and both the cumulative Fe dose received and the time elapsed since start of dialysis. This is an interesting observation that suggests that uremia itself may progressively lead to Fe accumulation in tissues over time.

Almost 40% of all hemodialysis patients in Australia have “unsafe” serum ferritin levels >500 µg/L (24). The perception of toxicity of therapeutic Fe in CKD patients is based largely on a limited number of observational studies reporting an increased infection rate in Fe-deficient subjects on Fe replacement (25) or in CKD patients with high ferritin concentrations (26) and increased CVD and death in dialysis patients (27,28). Serum ferritin was shown to correlate with LIC assessed noninvasively by a superconducting quantum interference device (17). Although our study is limited by small sample size and the selection of hemodialysis patients with elevated serum ferritin and variable duration of dialysis, it is consistent with previous postmortem studies showing a lack of association between excessive tissue Fe and high serum ferritin but a strong association with duration of dialysis for >3 years and a cumulative Fe dose of between 6 and 25 g (29). Our results do not corroborate the findings by Canavese et al. (17), and it is possible that differences in timing of the parenteral Fe administration in relation to the timing of assay for TSAT and ferritin (23) may account for the observed differences between studies. A lack of correlation between serum ferritin levels and LIC has also been reported in other diseases, such as the common liver disorder nonalcoholic fatty liver disease (19). Although discrepancies between LIC and ferritin levels could be influenced by the presence of overt or subclinical inflammatory states, this seems unlikely because the mean CRP in our study (4.9 ± 4.1 mg/L) did not differ from the mean CRP in the study of Canavese et al. (6.2 ± 8.3 mg/L). Proinflammatory cytokines such as IL-1β, IL-6, and TNF-α are known to increase the synthesis of ferritin through increased translation of ferritin mRNA (30,31). It is possible that higher amounts of ferritin may trap more body Fe and protect the individual against worsening infection, be-
cause free Fe is believed to enhance the formation of free oxygen radicals, which are the mediators of cell damage in infection-associated inflammation. Hence, inflammation-induced hyperferritinemia may result in a so-called “functional Fe deficiency,” which may be useful in “acute” inflammation by leading to anemia in subjects who already have significant comorbidity. The issue of how to treat Fe overload in this setting is problematic with no proven therapy. Improved knowledge of key regulators of Fe metabolism should result in further study and validation of new therapies for the treatment of Fe overload in CKD.

**References**


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